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		time-resol Anal Bioche	g human beta-s ved fluorescen m. 2003 Aug 1;3 2106 [PubMed -	ce. 19(1):49-55.	CE1) activity	using homog	geneous
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	□4:	<u>Luo KQ, Yu</u>	VC, Pu Y, Chang	gDC.		Related Arti	icles, Links
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	5 :	Takemoto K	, Nagai T, Miyaw	aki A, Miura M.	<u>.</u>	Related Arti	cles, Links
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	116:	<u>Onuki R, Na</u> <u>K.</u>	gasaki A, Kawasa	iki H, Baba T, U	Jyeda TQ, Taira	Related Arti	cles, Links
		significant Proc Natl Ac	ion by FRET in amyloid beta - ad Sci U S A. 200 9609 [PubMed - 1	-mediated cas 02 Nov 12;99(2	spase 8 activat 3):14716-21. Ep	cion.	

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	Application of the fluorescence resonance energy to studying the dynamics of caspase-3 activation during apoptosis in living HeLa cells. Biochem Biophys Res Commun. 2001 May 25;283(5):1054-66. PMID: 11355879 [PubMed - indexed for MEDLINE]	ng UV-induced
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	Quantitative analysis of fluorescent caspase substracells and identification of novel inhibitors of apopto Cell Death Differ. 2001 Jan;8(1):30-7. PMID: 11313700 [PubMed - indexed for MEDLINE]	
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	Development and application of a GFP-FRET intra for drug screening. J Biomol Screen. 2000 Oct;5(5):307-18. PMID: 11080689 [PubMed - indexed for MEDLINE]	cellular caspase assay
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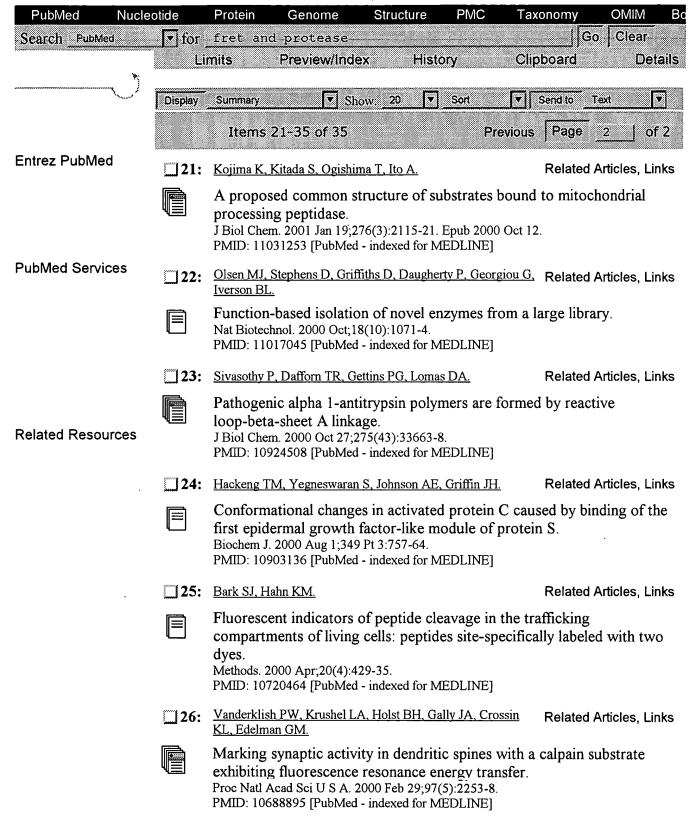
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L3 0 STEWARD/BI AND PROTEASE

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L4 1695 PROTEASE AND CLOSTRIDIUM

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L5 873 L4 AND ACTIVITY

=> s 15 and method

L6 142 L5 AND METHOD

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ANSWER 1 OF 27 WPIDS COPYRIGHT 2003 THOMSON DERWENT on STN

ACCESSION-NUMBER: 2003-278642 [27] WPIDS

DOC. NO. CPI:

C2003-072953

TITLE:

New purified peptide comprising a variant proaerolysin

amino acid sequence with a prostate-specific protease cleavage site and a functionally deleted

furin cleavage site, useful for treating prostate cancer.

B04 D16 DERWENT CLASS:

INVENTOR(S):

BUCKLEY, J T; DENMEADE, S R; ISAACS, J T

PATENT ASSIGNEE(S):

(UYJO) UNIV JOHNS HOPKINS; (UYVI-N) UNIV VICTORIA

INNOVATION & DEV CORP

COUNTRY COUNT:

101

PATENT INFORMATION:

PATENT NO KIND DATE WEEK LA PG

WO 2003018611 A2 20030306 (200327)* EN 83

RW: AT BE BG CH CY CZ DE DK EA EE ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SK SL SZ TR TZ UG ZM ZW

W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ OM PH PL PT RO RU SD SE SG SI SK SL TJ TM TN TR TT TZ UA UG US UZ VC VN YU ZA ZM ZW

APPLICATION DETAILS:

APPLICATION DATE PATENT NO KIND WO 2003018611 A2 WO 2002-US27061 20020823

PRIORITY APPLN. INFO: US 2001-314613P 20010824

2003-278642 [27] WPIDS

WO2003018611 A UPAB: 20030429

NOVELTY - A purified peptide comprising a variant proaerolysin amino acid sequence having a prostate-specific protease cleavage site and a functionally deleted furin cleavage site, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for:

- (1) treating prostate cancer by administering the peptide or the nucleic acid encoding the peptide to the subject, or contacting prostate cancer cells of the subject with the peptide;
- (2) systematically treating prostate cancer in a subject by removing prostate cancer cells from the subject, contacting the cells with the peptide to generate a cell lysate, and administering the cell lysate to the subject; a purified peptide comprising a variant proaerolysin amino acid sequence comprising a furin cleavage site, a functionally deleted proaerolysin binding domain, and a prostate-tissue specific binding domain;
- (3) a purified peptide comprising a variant Clostridium septicum alpha toxin amino acid sequence comprising a prostate-specific protease cleavage site and a functionally deleted furin cleavage site:
- (4) a purified peptide comprising a variant Bacillus thuringiensis delta-toxin amino acid sequence comprising a prostate-specific protease cleavage site and a functionally deleted wild type activation sequence;

- (5) a purified peptide comprising a variant human perforin amino acid sequence comprising prostate-specific **protease** cleavage site and a functionally deleted wild type perforin activation site; a purified peptide comprising a variant **Clostridium** septicum alpha **toxin** amino acid sequence comprising a furin cleavage site, a functionally deleted **Clostridium** septicum alpha **toxin** binding domain, and a prostate-tissue specific-binding domain;
- (6) a purified peptide comprising a variant Bacillus thuringiensis delta-toxin amino acid comprising wild type activation site, a functionally deleted Bacillus thuringiensis delta-toxin binding domain, and a prostate-tissue specific binding domain; a purified peptide comprising a variant human perforin amino acid sequence comprising a wild type activation site, a functionally deleted perforin binding domain, and a prostate-tissue specific binding domain; and
 - (7) a nucleic acid sequence encoding the peptide.

ACTIVITY - Cytostatic.

No biological data given.

MECHANISM OF ACTION - Peptide therapy.

USE - The peptides and **methods** are useful for treating prostate cancer (claimed).

Dwg.0/5

=> d 18 ibib abs 1-27

L8 ANSWER 1 OF 27 WPIDS COPYRIGHT 2003 THOMSON DERWENT on STN

ACCESSION NUMBER:

2003-278642 [27] WPIDS

DOC. NO. CPI:

C2003-072953

TITLE:

New purified peptide comprising a variant proaerolysin

amino acid sequence with a prostate-specific

protease cleavage site and a functionally deleted

furin cleavage site, useful for treating prostate cancer.

DERWENT CLASS:

INVENTOR(S):

BUCKLEY, J T; DENMEADE, S R; ISAACS, J T

PATENT ASSIGNEE(S):

(UYJO) UNIV JOHNS HOPKINS; (UYVI-N) UNIV VICTORIA

INNOVATION & DEV CORP

COUNTRY COUNT:

NT .

PATENT INFORMATION:

PATENT	ИО	KIND	DATE	WEEK	LΑ	PG
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B04 D16

101

WO 2003018611 A2 20030306 (200327)* EN 83

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W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ OM PH PL PT RO RU SD SE SG SI SK SL TJ TM TN TR TT TZ UA UG US UZ VC VN YU ZA ZM ZW

APPLICATION DETAILS:

PATENT NO	KIND	APPLICATION	DATE
WO 20030186	311 A2	WO 2002-US27061	20020823

PRIORITY APPLN. INFO: US 2001-314613P 20010824

AM 2003-270642 [27] WPIDS

AB W02003018611 A UPAB: 20030429

NOVELTY - A purified peptide comprising a variant proaerolysin amino acid sequence having a prostate-specific **protease** cleavage site and a

functionally deleted furin cleavage site, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for:

- (1) treating prostate cancer by administering the peptide or the nucleic acid encoding the peptide to the subject, or contacting prostate cancer cells of the subject with the peptide;
- (2) systematically treating prostate cancer in a subject by removing prostate cancer cells from the subject, contacting the cells with the peptide to generate a cell lysate, and administering the cell lysate to the subject; a purified peptide comprising a variant proaerolysin amino acid sequence comprising a furin cleavage site, a functionally deleted proaerolysin binding domain, and a prostate-tissue specific binding domain;
- (3) a purified peptide comprising a variant Clostridium septicum alpha toxin amino acid sequence comprising a prostate-specific protease cleavage site and a functionally deleted furin cleavage site;
- (4) a purified peptide comprising a variant Bacillus thuringiensis delta-toxin amino acid sequence comprising a prostate-specific protease cleavage site and a functionally deleted wild type activation sequence;
- (5) a purified peptide comprising a variant human perforin amino acid sequence comprising prostate-specific protease cleavage site and a functionally deleted wild type perforin activation site; a purified peptide comprising a variant Clostridium septicum alpha toxin amino acid sequence comprising a furin cleavage site, a functionally deleted Clostridium septicum alpha toxin binding domain, and a prostate-tissue specific binding domain;
- (6) a purified peptide comprising a variant Bacillus thuringiensis delta-toxin amino acid comprising wild type activation site, a functionally deleted Bacillus thuringiensis delta-toxin binding domain, and a prostate-tissue specific binding domain; a purified peptide comprising a variant human perforin amino acid sequence comprising a wild type activation site, a functionally deleted perforin binding domain, and a prostate-tissue specific binding domain; and
 - (7) a nucleic acid sequence encoding the peptide.

ACTIVITY - Cytostatic.

No biological data given.

MECHANISM OF ACTION - Peptide therapy.

USE - The peptides and methods are useful for treating prostate cancer (claimed). Dwg.0/5

ANSWER 2 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: DOCUMENT NUMBER:

2003:113075 BIOSIS PREV200300113075

TITLE:

Fluorigenic substrates for the protease

activities of botulinum neurotoxins, serotypes A,

B, and F.

AUTHOR(S):

Schmidt, James J. (1); Stafford, Robert G.

CORPORATE SOURCE:

(1) Toxinology and Aerobiology Division, United States Army Medical Research Institute of Infectious Diseases, 1425

Porter St., Fort Detrick, MD, 21702-5011, USA:

james.schmidt@det.amedd.army.mil USA

SOURCE:

Applied and Environmental Microbiology, (January 2003,

2003) Vol. 69, No. 1, pp. 297-303. print.

ISSN: 0099-2240.

DOCUMENT TYPE:

Article English

LANGUAGE:

The seven botulinum neurotoxins (BoNTs) are zinc metalloproteases that cleave neuronal proteins involved in neurotransmitter release and are among the most toxic natural products known. High-throughput BoNT assays are needed for use in antibotulinum drug discovery and to characterize

BoNT protease activities. Compared to other proteases, BoNTs exhibit unusually stringent substrate requirements with respect to amino acid sequences and polypeptide lengths. Nonetheless, we have devised a strategy for development of fluorigenic BoNT protease assays, based on earlier structure-function studies, that has proven successful for three of the seven serotypes: A, B, and F. In synthetic peptide substrates, the P1 and P3' residues were substituted with 2,4-dinitrophenyl-lysine and S-(N-(4-methyl-7dimethylamino-coumarin-3-yl)-carboxamidomethyl)-cysteine, respectively. By monitoring the BoNT-catalyzed increase in fluorescence over time, initial hydrolysis rates could be obtained in 1 to 2 min when BoNT concentrations were 60 ng/ml (about 1 nM) or higher. Each BoNT cleaved its fluorigenic substrate at the same location as in the neuronal target protein, and kinetic constants indicated that the substrates were selective and efficient. The fluorigenic assay for BoNT B was used to characterize a new competitive inhibitor of BoNT B protease activity with a Ki value of 4 muM. In addition to real-time activity measurements, toxin concentration determinations, and kinetic studies, the BoNT substrates described herein may be directly incorporated into automated high-throughput assay systems to screen large numbers of compounds for potential antibotulinum drugs.

ANSWER 3 OF 27 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 1

ACCESSION NUMBER:

2002:353597 CAPLUS

DOCUMENT NUMBER:

136:365216

TITLE:

Recombinant light chains of botulinum neurotoxins and

light chain fusion proteins for use in research and

clinical therapy

INVENTOR(S):

Smith, Leonard A.; Jensen, Melody

PATENT ASSIGNEE(S):

United States Army Medical Research and Material

Command, USA

SOURCE:

PCT Int. Appl., 166 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

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PATENT NO.
                 KIND DATE
                                         APPLICATION NO. DATE
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                                    WO 2001-US47230 20011106
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            GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,
            LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH,
            PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ,
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                      Α5
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                                                         20011106
PRIORITY APPLN. INFO.:
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                                      US 1999-133866P P 19990512
                                      US 1999-133867P P 19990512
                                      US 1999-133868P P 19990512
                                      US 1999-133869P P 19990512
                                      US 1999-133873P P 19990512
                                      US 1999-146192P P 19990729
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WO 2000-US12890 W 20000512 US 2000-611419 A1 20000706 WO 2001-US47230 W 20011106

Botulinum neurotoxins, the most potent of all toxins, induce AΒ lethal neuromuscular paralysis by inhibiting exocytosis at the neuromuscular junction. The light chains (LC) of these dichain neurotoxins are a new class of zinc-endopeptidases-that specifically cleave the synaptosomal proteins, SNAP-25, VAMP, or syntaxin at discrete sites. The present invention relates to the construction, expression, purifn., and use of synthetic or recombinant botulinum neurotoxin genes. For example, a synthetic gene for the LC of the botulinum neurotoxin serotype A (BoNT/A) was constructed and overexpressed in Escherichia coli. The gene product was purified from inclusion bodies. The methods of the invention can provide 1.1 g of the LC per L of culture. The LC product was stable in soln. at 4.degree. for at least 6 mo. This rBoNT/A LC was proteolytically active, specifically cleaving the Glu-Arg bond in a 17-residue synthetic peptide of SNAP-25, the reported cleavage site of BoNT/A. Its calcd. catalytic efficiency kcat/Km was higher than that reported for the native BoNT/A dichain. Treating the rBoNT/A LC with mercuric compds. completely abolished its activity, most probably by modifying the cysteine-164 residue located in the vicinity of the active site. About 70% activity of the LC was restored by adding Zn2+-free, apo-LC prepn. The LC was nontoxic to mice and failed to elicit neutralizing epitope(s) when the animals were vaccinated with this protein. In addn., injecting rBoNT/A LC into sea urchin eggs inhibited exocytosis-dependent plasma membrane resealing.

ANSWER 4 OF 27 WPIDS COPYRIGHT 2003 THOMSON DERWENT on STN

ACCESSION NUMBER: 2002-590661 [63] WPTDS

DOC. NO. CPI: TITLE:

C2002-167115

Methods for surface display of proteins, e.g.

displaying a protein on the surface of spores, improving a protein or isolating a substance, by transforming a host cell harboring a genetic carrier with spore or virus

with the vector library.

DERWENT CLASS:

INVENTOR(S):

B04 C06 D16 BAN, J G; CHOI, S G; JUNG, H C; CHOI, S K; PAN, J G

PATENT ASSIGNEE(S):

COUNTRY COUNT:

(GENO-N) GENOFOCUS CO LTD 96

PATENT INFORMATION:

PATENT NO KIND DATE WEEK LΑ

WO 2002055561 A1 20020718 (200263)* EN 118

RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZM ZW

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KR 2002061218 A 20020724 (200305)

APPLICATION DETAILS:

PATENT NO	KIND	APPLICATION	DATE
WO 20020555	61 Al	WO 2002-KR59	20020115
KR 200206123	18 A	KR 2001-2156	20010115

PRIORITY APPLN. INFO: KR 2001-2156 20010115

AN 2002-590661 [63] WPIDS

NOVELTY - Methods for preparing a protein surface-displayed on a genetic carrier, improving a protein or isolating a substance in a mixture comprising transforming a host cell harboring a genetic carrier consisting of a spore or virus with the vector library, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are included for:

- (1) preparing a protein surface displayed on a genetic carrier comprising:
- (a) transforming a host cell harboring the genetic carrier consisting of spore or virus with a vector containing a gene encoding the protein;
- (b) culturing the transformed host cell and expressing the protein in the host cell; and
- (c) allowing to form non-covalent bonds between the expressed protein and a surface of the genetic carrier so that the protein is displayed on the surface of the genetic carrier;
 - (2) improving a protein comprising:
- (a) constructing a gene library of the protein by means of mutating the gene encoding the protein;
- (b) preparing a vector library containing the constructed gene library;
 - (c) employing step (1-a) and (1-b) above;
- (d) obtaining a genetic carrier library by means of allowing to form noncovalent bonds between the expressed protein variant and a surface of the genetic carrier so that the variant is displayed on the surface of the genetic carrier; and
- (e) screening the genetic carrier displaying on its surface the variant of the protein having a desired property;
 - (3) isolating a substance in mixture comprising:
- (a) constructing a gene library encoding a variant of binding protein or its binding domain by means of mutating the gene encoding the binding protein or binding domain as protein of interest;
 - (b) employing (2-b) or (1-a);
- (c) obtaining a genetic carrier library by means of allowing to form noncovalent bonds between the expressed binding protein variant or binding domain and a surface of the genetic carrier so that the variant is displayed on the surface of the genetic carrier;
- (d) contacting the genetic carrier library with a predetermined substance and screening an improved binding protein or its binding domain by means of selecting the genetic carrier displaying on its surface the variant binding the predetermined substance; and
- (e) contacting the genetic carrier displaying on its surface the improved binding protein or its binding domain with the mixture to isolate the substance in mixture;
- (4) a vector for displaying on the surface of a genetic carrier a protein, which comprises a replication origin, an antibiotic-resistance gene, a restriction site, or a gene encoding the protein, where the protein, when expressed in a host cell, is capable of forming noncovalent bond to the surface of genetic carrier;
- (5) a microbial transformant, which is produced by transforming a host cell harboring spores or viruses with the vector;
- (6) a complex between genetic carrier and protein, characterized in that the complex is prepared by displaying on the surface of the genetic carrier, hormone, hormone analogue, enzyme, enzyme inhibitor, signal transduction protein or its fragment, antibody or its fragment, single chain antibody, binding protein or its fragment, peptide, antigen, adhesive protein, structural protein, regulatory protein, toxin protein, cytokine, transcription regulatory protein, blood clotting protein or plant defense-inducing protein;
- (7) a genetic carrier library displaying on its surface variants of a protein, prepared by a process comprising the steps (2-a) (2-e);
- (8) bioconversion using protein with activity for conversion reaction, characterized in that the method employs

the complex between genetic carrier and protein;

- (9) producing an antibody to an antigen in vertebrates, characterized in that the method comprises administering to vertebrates a composition containing an immunological amount of the complex between genetic carrier and protein; and
- (10) a protein microarray comprising a solid substrate and a material immobilized onto the substrate, characterized in that the material immobilized onto the substrate is selected from the complex between genetic carrier and protein; and the genetic carrier library.

USE - The method is useful for surface display of proteins, particularly for displaying a protein on the surface of e.g. spores, for improving a protein, or for isolating a substance. These methods are useful in obtaining monoclonal variants from a large library, or in high-throughput screening of antibodies for use in therapy (e.g. as vaccines), diagnosis or analysis. Dwg.0/8

ANSWER 5 OF 27 WPIDS COPYRIGHT 2003 THOMSON DERWENT on STN 1.8

ACCESSION NUMBER: 2002-195744 [25] WPIDS

DOC. NO. CPI:

C2002-060478

TITLE:

Novel synthetic peptides which include cell-growth affecting peptides and peptides which enhance or inhibit cellular protein production, useful for enhancing or inhibiting cell growth or cellular protein production.

DERWENT CLASS:

B04 D16

INVENTOR(S):

CAMPBELL, R L; ERICKSON, B W; HAALAND, P D; LLOYD, S A;

SHERMAN, D B; STEWART, W W

PATENT ASSIGNEE(S):

(BECT) BECTON DICKINSON & CO

COUNTRY COUNT:

31

PATENT INFORMATION:

PATENT NO	KIND DATE	WEEK	LΑ	PG
				

WO 2002002591 A2 20020110 (200225)* EN 41

RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE TR

W: AU BR CA CN ES IL JP KR MX NZ RU SG

AU 2001075173 A 20020114 (200237)

A2 20030611 (200339) EN EP 1317475

R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE TR

APPLICATION DETAILS:

PATENT NO K	IND	AP.	PLICATION	DATE
WO 2002002591 AU 2001075173 EP 1317475		AU EP	2001-75173 2001-941853	20010604 20010604 20010604 20010604

FILING DETAILS:

PΑ	TENT NO	KIND			PA7	CENT	NO
AU	200107517	73 A	Based	on	WO	2002	002591
EF	1317475	A2	Based	on	WO	2002	002591

PRIORITY APPLN. INFO: US 2000-608892 20000630

AN 2002-195744 [25] WPIDS

ΔB WO 200202591 A UPAB: 20020418

> NOVELTY - Synthetic peptides (I) which include cell-growth affecting peptides and peptides which enhance or inhibit cellular protein production, selected from 94 peptides given in the specification, are new.

DETAILED DESCRIPTION - (I) is selected from 94 peptides given in the specification such as GEAL, KLAL, EKAL, ESAL, NDAL, NNAL, SNAL, VNAL, KKAL or SKKA.

INDEPENDENT CLAIMS are also included for the following:

- (1) a peptide library (II) comprising chemically synthesized peptides, each of the peptides comprising an N-terminal or C-terminal amino acid associated with enzymatic or chemical cleavage of a polypeptide and one or more additional amino acids;
 - (2) a peptide (III) selected from (II);
 - (3) a cell or tissue culture medium (IV) comprising (I) or (III); and
 - (4) production of (III).

ACTIVITY - Antibacterial.

No supporting data provided.

MECHANISM OF ACTION - Inhibitor/enhancer of cellular protein production and/or cellular growth.

No supporting data provided.

USE - (I) or (III) is useful for enhancing or inhibiting cell growth of **Clostridium** perfringens or cellular protein production of beta -toxin, by culturing the cells or tissues in the presence of about 0.1-25 mM, preferably 1.0-12 mM of (I) or (III) (claimed). (II) is useful for rapid identification of biologically active compounds which affect the properties of cells in culture. (II) or (III) is useful in concatemer-based recombinant expression methods or in large-scale, economical recombinant production methods.

ADVANTAGE - (I) reduces the number and quantity of undefined components in culture media, reduces the need for animal-derived components, improves media consistency and quality control, and provides a method for precisely controlling and adjusting performance of the cell culture.

Dwg.0/0

L8 ANSWER 6 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER:

2002:168833 BIOSIS PREV200200168833

DOCUMENT NUMBER: TITLE:

In vitro reconstitution of the Clostridium

botulinum type D progenitor toxin.

AUTHOR(S):

SOURCE:

Kouguchi, Hirokazu; Watanabe, Toshihiro; Sagane, Yoshimasa;

Sunagawa, Hiroyuki; Ohyama, Tohru (1)

CORPORATE SOURCE:

(1) Department of Food Science and Technology, Faculty of Bioindustry, Tokyo University of Agriculture, 196 Yasaka, Abashiri, 099-2493: t-oyama@bioindustry.nodai.ac.jp Japan Journal of Biological Chemistry, (January 25, 2002) Vol. 277, No. 4, pp. 2650-2656. http://www.jbc.org/. print.

ISSN: 0021-9258.

DOCUMENT TYPE:

Article English

LANGUAGE: Clostridium botulinum type D strain 4947 produces two different AB sizes of progenitor toxins (M and L) as intact forms without proteolytic processing. The M toxin is composed of neurotoxin (NT) and nontoxic-nonhemagglutinin (NTNHA), whereas the L toxin is composed of the M toxin and hemagglutinin (HA) subcomponents (HA-70, HA-17, and HA-33). The HA-70 subcomponent and the HA-33/17 complex were isolated from the L toxin to near homogeneity by chromatography in the presence of denaturing agents. We were able to demonstrate, for the first time, in vitro reconstitution of the L toxin formed by mixing purified M toxin, HA-70, and HA-33/17. The properties of reconstituted and native L toxins are indistinguishable with respect to their gel filtration profiles. native-PACE profiles, hemagglutination activity, binding activity to erythrocytes, and oral toxicity to mice. M toxin, which contained nicked NTNHA prepared by treatment with trypsin, could no longer be reconstituted to the L toxin with HA

subcomponents, whereas the L toxin treated with proteases was not degraded into M toxin and HA subcomponents. We conclude that the M toxin forms first by assembly of NT with NTNHA and is subsequently converted to the L toxin by assembly with HA-70 and H-33/17.

L8 ANSWER 7 OF 27 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 2

ACCESSION NUMBER:

2003:277358 CAPLUS

TITLE:

In vitro determination of specific toxicity in tetanus

vaccines

AUTHOR(S):

Kegel, B.; Bonifas, U.; Silberbach, K.; Kramer, B.;

Weisser, K.

CORPORATE SOURCE:

Federal Agency for Sera and Vaccines, Paul-Ehrlich-Institute, Langen, Germany

SOURCE:

Developments in Biologicals (Basel, Switzerland) (2002), 111 (Advancing Science and Elimination of the Use of Laboratory Animals for Development and Control

of Vaccines and Hormones), 27-33 CODEN: DBEIAI; ISSN: 1424-6074

PUBLISHER:

S. Karger AG

DOCUMENT TYPE:

Journal English

LANGUAGE:

AΒ

Tetanus vaccine is prepd. from detoxified tetanus neurotoxin. To ensure the absence of residual toxin activity or to exclude

the reversion to toxicity reliable control testing is based on in vivo methods, because no in vitro assay provides the required specificity and sensitivity. Tetanus neurotoxin is a 150 kDa protein produced by Clostridium tetani. The 50 kDa light chain of this neurotoxin belongs to the family of zinc metalloproteases. It cleaves synaptobrevin, a small synaptic vesicle protein, which is involved in neuroexocytosis, at the single Q76-F77 peptide bond. To develop a sensitive in vitro assay capable of quantifying the proteolytic

activity of this toxin, we used as substrate a recombinant fragment of synaptobrevin2 (1-97). For detecting the cleavage products a peptide antibody raised against the N-terminal cleavage site was used. In Western Blot anal. only the cleaved substrate was detected while the uncleaved substrate showed no signal. In different approaches, recombinant synaptobrevin was either. (i) bound to a microtitre plate, reduced toxin was added and the N-terminal cleavage product was detected by a specific antibody or. (ii) the cleavage was performed in test tubes, the samples were transferred to a microtitre plate and immobilized cleavage products were detected. When toxoid or crude toxin is used, non-specific cleavage of synaptobrevin substrate occurs. Depending on the toxoid used different patterns of degrdn. of substrate are visible in Western Blots. Different protease inhibitors and reaction conditions seem to have an effect on the inhibition of this non-specific cleavage.

REFERENCE COUNT:

THERE ARE 5 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L8 ANSWER 8 OF 27 CAPLUS COPYRIGHT 2003 ACS on STN

5

ACCESSION NUMBER:

2001:152855 CAPLUS

DOCUMENT NUMBER:

134:203683

TITLE:

Recombinant construction and expression of

single-chain activatable neurotoxins

INVENTOR(S):

Dolly, J. Oliver; Li, Yan; Chan, Kuo Chion

PATENT ASSIGNEE(S):

Allergan Sales, Inc., USA

SOURCE:

PCT Int. Appl., 90 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

T: 1

SOURCE:

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APPLICATION NO. DATE
                  KIND DATE
     PATENT NO.
     WO 2001014570 A1 20010301 WO 2000-US23427 20000825
        W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN,
            CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR,
            HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT,
            LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU,
             SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN,
            YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
         RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY,
            DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ,
             CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
                    A 20020402 BR 2000-12759 20000825
A1 20020522 EP 2000-964920 20000825
     BR 2000012759
     EP 1206554
           AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
             IE, SI, LT, LV, FI, RO, MK, CY, AL
                                          JP 2001-518882 20000825
     JP 2003507073
                     T2 20030225
                                       US 1999-150710P P 19990825
PRIORITY APPLN. INFO.:
                                       WO 2000-US23427 W 20000825
     Compns. comprising activable recombinant neurotoxins and polypeptides
AB
     derived therefrom. The invention also comprises nucleic acids encoding
     such polypeptides, and methods of making such polypeptides and
     nucleic acids. Thus, a single-chain protein is constructed by genetic
     engineering techniques comprising the functional domains of a clostridial
     neurotoxin H chain and some or all of the functions of a clostridial
     neurotoxin L chain, and having an inserted proteolytic cleavage site
     located between the H domain and the L domain by which the single-chain
     protein may be cleaved to produce the individual chains, prefereably
     covalently linked by a disulfide linkage. To minimize the safety risk
     assocd. with handling neurotoxins, they are expressed as their low
     activity (or inactive) single-chian proforms, and then carefully
     activated via cleavage at a site designed to have a high degree of
     specificity to proteolytic enzymes which do not normally occur in humans.
     The interchain loop region of the Clostridium botulinum subtype
     E neurotoxin, which is normally resistant to proteolytic nicking in the
     bacterium and mammals, is modified to include the inserted proteolytic
     cleavage site. Single-chain tetanus toxins contg. a bovine
     enterokinase cleavage site are expressed from Escherichia coli and shown
     to induce in vitro paralysis using the mouse phrenic nerve hemi-diaphragm
     assay. Further modification of single-chain tetanus toxin to
     remove proteolytic cleavage sites reduces the toxicity of unnicked
     recombinant toxin. Single-chain botulin type A and E
     neurotoxins are also described.
                               THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS
REFERENCE COUNT:
                               RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT
     ANSWER 9 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN
L8
ACCESSION NUMBER: 2001:283768 BIOSIS
                   PREV200100283768
DOCUMENT NUMBER:
TITLE:
                   High-throughput fluorogenic assay for determination of
                   botulinum type B neurotoxin protease
                   activity.
                   Anne, Christine; Cornille, Fabrice; Lenoir, Christine;
AUTHOR(S):
                   Roques, Bernard P. (1)
CORPORATE SOURCE:
                   (1) Departement de Pharmacochimie Moleculaire et
                   Structurale, U266 INSERM, UMR 8600 CNRS, UFR des Sciences
                   Fharmaceutiques et Biologiques, 4, Avenue de
                   l'Observatoire, 75270, Paris Cedex 06:
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roques@pharmacie.univ-paris5.fr France

Analytical Biochemistry, (April 15, 2001) Vol. 291, No. 2,

pp. 253-261. print. ISSN: 0003-2697.

DOCUMENT TYPE:

Article English English

LANGUAGE: SUMMARY LANGUAGE:

Botulinum neurotoxins are responsible for botulism, a flaccid muscular paralysis caused by inhibition of acetylcholine release at the neuromuscular junction. This occurs by cleavage of conserved proteins involved in exocytosis such as synaptobrevin by the zinc metallopeptidase activity of the light chain of some botulinum neurotoxins. Botulism, for which there is presently no therapy available, is a relatively widespread disease that may result in death. Consequently, the development of drugs able to inhibit the hydrolytic activity of these neurotoxins is of great interest. Design and screening of such inhibitors could be largely facilitated by using high-throughput assays. With this aim, a novel in vitro test for quantifying the proteolytic activity of botulinum type B neurotoxin was developed. The substrate is the 60-94 fragment of human synaptobrevin-1 which was modified by introduction of the fluorescent amino acid L-pyrenylalanine in position 74 and a p-nitrophenylalanyl residue as quenching group in position 77. The cleavage of Syb 60-94 (Pya74, Nop77) by the toxin active chain occurs selectively between residues 76 and 77 as in the case of the unmodified synaptobrevin and is directly quantified by measuring the strong fluorescence of the formed metabolite Syb 60-76 (Pya74). This is the easiest, quickest, and cheapest assay described to date for measuring the proteolytic activity of botulinum type B neurotoxin. It can be easily automated for high-throughput screening. Moreover, amounts of about 3.5 pg/ml of botulinum type B neurotoxin could be detected by this method.

ANSWER 10 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: DOCUMENT NUMBER:

2000:111428 BIOSIS PREV200000111428

TITLE:

Rescue of exocytosis in botulinum toxin

A-poisoned chromaffin cells by expression of

cleavage-resistant SNAP-25: Identification of the minimal

essential C-terminal residues.

AUTHOR(S):

O'Sullivan, Gregory A.; Mohammed, Nadiem; Foran, Patrick

G.; Lawrence, Gary W.; Dolly, J. Oliver (1)

CORPORATE SOURCE:

(1) Department of Biochemistry, Imperial College, London,

SW7 2AY UK

SOURCE:

Journal of Biological Chemistry, (Dec. 24, 1999) Vol. 274,

No. 52, pp. 36897-36904.

ISSN: 0021-9258.

DOCUMENT TYPE: LANGUAGE:

Article English SUMMARY LANGUAGE: English

Botulinum neurotoxin (BoNT) types A and B selectively block exocytosis by cleavage of SNAP-25 and synaptobrevin, respectively; in humans, many months are required for full recovery from the resultant neuromuscular paralysis. To decipher the molecular basis for such prolonged poisoning, intoxication in adreno-chromaffin cells was monitored over 2 months. Exocytosis from BoNT/B-treated cells resumed after 56 days because of the appearance of intact synaptobrevin. However, inhibition continued in BoNT/A-treated cells, throughout the same interval, with a continued predominance of cleaved SNAP-25-(1-197) over the intact protein. When recovery from poisoning was attempted by transfection of the latter cells with the gene encoding full-length SNAP-25-(1-206), no restoration of cmocytosis ensued even after 3 weeks. To ascertain if this failure was because of the persistence of the toxin's protease activity, the cells were transfected with BoNT/A-resistant SNAP-25 constructs; importantly, exocytosis was rescued. C-terminal truncation of

the toxin-insensitive SNAP-25 revealed that residues 1-201, 1-202, 1-203 afforded a significant return of exocytosis, unlike shorter forms 1-197, -198, -199, or -200; accordingly, mutants M202A or L203A of full-length SNAP-25 rescued secretion. These findings give insights into the C-terminal functional domain of SNAP-25, demonstrate the longevity of BoNT/A protease, and provide the prospect of a therapy for botulism.

ANSWER 11 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1999:186708 BIOSIS

DOCUMENT NUMBER:

PREV199900186708

TITLE:

Molecular properties of a hemagglutinin purified from type

a Clostridium botulinum.

AUTHOR(S):

Sharma, Shashi Kant; Fu, Fen-Ni; Singh, Bal Ram (1)

CORPORATE SOURCE:

(1) Department of Chemistry and Biochemistry, University of Massachusetts Dartmouth, North Dartmouth, MA, 02747 USA

SOURCE:

Journal of Protein Chemistry, (Jan., 1999) Vol. 18, No. 1,

pp. 29-38.

ISSN: 0277-8033.

DOCUMENT TYPE:

Article English

LANGUAGE:

AΒ Clostridium botulinum causes the food poisoning disease botulism by producing botulinum neurotoxin, the most potent toxin known. The neurotoxin is produced along with a group of neurotoxin-associated proteins, or NAPs, which protect it from the low pH and proteases of the gastrointestinal tract. Recently, we isolated one of the major components of NAPs, a 33-kDa hemagglutinin (Hn-33) (Fu et al. (1998), J. Protein Chem. 17, 53-60). In this study, we present molecular properties of Hn-33 derived from several biochemical and biophysical techniques. Hn-33 in pure form requires a 66-fold lower concentration of sugar inhibition of its hemagglutination activity than in its complexed form with the neurotoxin and other NAPs. However, its protease resistance is not affected by sugar binding. Based on FT-IR and circular dichroism (CD) analysis, Hn-33 is a predominantly beta-sheet protein (74-77%). Hn-33 analysis by laser desorption mass spectrometry and size exclusion column chromatography reveals that it exists predominantly in a dimeric form in the aqueous solution. Even a very low concentration of SDS (0.05%) irreversibly destroyed the biological activity of Hn-33 by changing its secondary structure as revealed by far-UV CD analysis.

ANSWER 12 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: DOCUMENT NUMBER:

1997:440312 BIOSIS PREV199799739515

TITLE:

The propeptide of Clostridium septicum alpha

toxin functions as an intramolecular chaperone and is a potent inhibitor of alpha toxin-dependent

cytolysis.

AUTHOR(S):

Sellman, Bret R.; Tweten, Rodney K. (1)

CORPORATE SOURCE:

(1) Microbiol. Immunol., Univ. Oklahoma Health Sci. Cent.,

Oklahoma City, OK 73190 USA

SOURCE:

Molecular Microbiology, (1997) Vol. 25, No. 3, pp. 429-440.

ISSN: 0950-382X.

DOCUMENT TYPE:

Article

LANGUAGE:

English

Clostridium septicum alpha toxin is activated by a proteolytic cleavage at Arg-398 in its carboxy terminus, which yields a 41.3-kDa cytolytically active toxin and a 5.1-kDa propeptide. Studies were performed to determine when the propeptide dissociated from the toxin after proteolytic activation of the protoxin (AT-pro) and to demonstrate the chaperone activity of the propeptide. The propeptide was found to remain associated with the toxin after

activation with trypsin (AT-act) when analysed by gel filtration or affinity chromatography of a polyhistidine-tagged derivative that contained the polyhistidine tag on the propeptide. The affinity of the propeptide for the toxin was decreased significantly when a mutation was introduced in which Val-400 was converted to a cysteine residue. This mutation destabilized the interaction of the propeptide with the toxin and the propeptide was found to dissociate from the toxin under the same gel-filtration conditions used for the wild-type toxin. The separation of the propeptide in the V400C mutant did not affect the cytolytic activity of the toxin and therefore the propeptide was not necessary for cytolytic activity. These data suggested that the propeptide did not dissociate from the main body of the toxin after proteolysis. Further analysis demonstrated that purified propeptide was a potent inhibitor of alpha toxin activity, which inhibited the oligomerization of alpha toxin into a functional pore. These data suggest that the propeptide does not participate in the final oligomerized complex and that oligomerization appears to displace the propeptide from AT-act. The importance of the propeptide to the solution stability of alpha toxin was also demonstrated. When AT-pro was activated in solution with trypsin a significant level (approximately 50%) of inactive aggregate formed. This aggregate, which could be removed by centrifugation at $14\ 000\ \text{times}$ g, was made up of both SDS-sensitive and -resistant aggregates, suggesting that a variety of inactive aggregates formed when the monomers interacted in solution. Significantly higher levels of haemolytic activity (approximately 16-fold) were observed when alpha toxin was proteolytically activated after membrane binding instead of in solution. These results support the role of the propeptide as an intramolecular chaperone that stabilizes the monomeric AT-pro and shuttles it to the membrane where it is activated by protease, oligomerizes into a pre-pore complex and forms a pore. The data suggest that oligomerization of the toxin displaces the propeptide from the monomer form of alpha toxin and that the propeptide does not participate in, and is not necessary to, the final cytolytic complex.

L8 ANSWER 13 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1997:410119 BIOSIS DOCUMENT NUMBER: PREV199799702162

TITLE: A biotinylated perfringolysin O derivative: A new probe for

detection of cell surface cholesterol.

AUTHOR(S): Iwamoto, Machiko (1); Morita, Ikuo; Fukuda, Mitsugu;

Murota, Sei-Itsu; Ando, Susumu; Ohno-Iwashita, Yoshiko

CORPORATE SOURCE: (1) Dep. Membrane Biochem., Tokyo Metropolitan Inst.

Gerontol., 35-2 Sakae-cho, Itabashi-ku, Tokyo 173 Japan

SOURCE: Biochimica et Biophysica Acta, (1997) Vol. 1327, No. 2, pp.

222-230.

ISSN: 0006-3002.

DOCUMENT TYPE: Article LANGUAGE: English

theta-Toxin is a cholesterol-binding, pore-forming cytolysin of Clostridium perfringens. To detect cell surface cholesterol, we prepared a theta-toxin derivative, BC-theta by biotinylation of a protease-nicked theta-toxin, which has the same binding affinity for cholesterol as theta-toxin without cytolytic activity. Human erythrocytes, V79 cells and human umbilical vein endothelial cells (HUVEC), were stained with BCO coupled with FITC-avidin, and then the cells were analyzed by either flow cytometry or laser confocal microscopy. The fluorescence intensity increased in both intact and briefly fixed cells when treated with BC-theta. BC-theta-treated V79 cells were stained by neither trypan blue nor propidium iodide, indicating that BC theta stained just the outer

surface of the plasma membrane of vital cells. Treatment of the cells with digitonin, a cholesterol sequestering reagent, decreased the fluorescence intensity to the background level, indicating that BC-theta staining is specific for cholesterol. The fluorescence intensity of erythrocytes pre-permeabilized with a small amount of theta-toxin increased more than ten-fold, suggesting higher cholesterol contents in the inner layer of the plasma membrane. When cells were cultured with cholesterol-depleted medium, the fluorescence intensity stained by BC-theta decreased remarkably in V79 cells, but did not change in HUVEC. This indicates that cell surface cholesterol may be provided in different ways with these two cell lines. These results suggest that BC theta can be a useful probe for visualizing cell surface cholesterol and for evaluating the effects of cellular events on the topology and distribution of cholesterol.

L8 ANSWER 14 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1996:569359 BIOSIS DOCUMENT NUMBER: PREV199799298715

TITLE: Contribution of individual tryptophan residues to the

structure and activity of THETA-toxin

(perfringolysin O), a cholesterol-binding cytolysin.

AUTHOR(S): Sekino-Suzuki, Naoko; Nakamura, Megumi; Mitsui, Ken-Ichiro;

Ohno-Iwashita, Yoshiko (1)

CORPORATE SOURCE: (1) Dep. Enzyme Biochemistry, Tokyo Metropolitan Inst.

Gerontol., Sakae-cho, Itabashi-ku, Tokyo 173 Japan

SOURCE: European Journal of Biochemistry, (1996) Vol. 241, No. 3,

pp. 941-947. ISSN: 0014-2956.

DOCUMENT TYPE: Article LANGUAGE: English

theta-Toxin (perfringolysin O), secreted by Clostridium perfringens, shares with other known thiol-activated toxins a conserved undecapeptide, ECTGLAWEWWR, located in the C-terminal region of the protein and containing the unique cysteine of the molecule. Single and double amino acid substitutions were created in the theta-toxin molecule to investigate the role of individual tryptophan residues in the lytic activity of theta-toxin. Wild-type and mutant theta-toxins were overproduced in Escherichia coli by means of a T7 RNA polymerase/promoter system and purified. The relative hemolytic activities of four mutant toxins, each with a Trp to Phe substitution outside the common Cys-containing region, were more than 60% that of wild-type theta-toxin. In contrast, mutant toxins with Phe replacements within the Cys-containing region (at Trp436, Trp438 or Trp439) showed significantly reduced hemolytic and erythrocyte-membrane binding activities. The largest reduction in binding affinity, more than 100-fold, was observed for Trp438 mutant toxins. However, the mutants retain binding specificity for cholesterol and the ability to form are shaped and ring-shaped structures on membranes. These results indicate that the low hemolytic activities of these mutant toxins can be ascribed, at least in part, to reduced binding activities. With respect to protease susceptibility and far-ultraviolet circular-dichroism spectra, only the W436 fwdarw F mutant toxin, showed any considerable difference from wild-type toxin in secondary or higher-order structures, indicating that Trp436 is essential for maintenance of toxin structure.

L8 ANSWER 15 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STM

DUPLICATE 3

ACCESSION NUMBER: 1996:422530 BIOSIS DOCUMENT NUMBER: PREV199699153586

TITLE: Nitric oxide inhibits rat intestinal secretion by

Clostridium difficile Toxin A but not

Vibrio cholerae enterotoxin.

AUTHOR(S): Qiu, Bosheng; Pothoulakis, Charalabos; Nikulasson, Ignazio

Castagliuolo Zigfus; Lamont, J. Thomas (1)

CORPORATE SOURCE: (1) Div. Gastroenterol., Beth Israel Hosp., 330 Brookline

Ave., Boston, MA 02215 USA

SOURCE: Gastroenterology, (1996) Vol. 111, No. 2, pp. 409-418.

ISSN: 0016-5085.

DOCUMENT TYPE: Article LANGUAGE: English

Background & Aims: Intestinal inflammation is associated with increased synthesis of nitric oxide, whereas inhibition of NO synthase (NOS) reduces experimental chronic intestinal inflammation. The aim of this study was to test the effects of NO blockers and donors on acute intestinal inflammation induced by Clostridium difficile toxin A in rat ileum. Methods: Rats received NOS inhibitors or NO donors before measurement of toxin-mediated ileal secretion and permeability changes. Mucosal mast cell and neutrophil activity were measured by release of rat mast cell protease II and myeloperoxidase activity, respectively. Results: NOS inhibitors augmented but an NO donor inhibited toxin A-mediated ileal secretion and permeability when given before but not after toxin administration. Neither an NOS inhibitor nor an NO donor had any effect on cholera toxin-mediated secretion. Mast cell degranulation and neutrophil infiltration occurred after injection of toxin A or an NOS inhibitor, whereas the NO donor blocked both toxin A effects. Conclusions: NOS inhibitors augmented and an NO donor blocked the intestinal effects of toxin A but not of cholera toxin . NO protects against toxin A by inhibition of intestinal mast cells and neutrophils, which are activated by toxin A, but not by cholera toxin.

L8 ANSWER 16 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1995:117495 BIOSIS DOCUMENT NUMBER: PREV199598131795

TITLE: Differences in the Protease Activities

of Tetanus and Botulinum B Toxins Revealed by the

Cleavage of Vesicle-Associated Membrane Protein and Various

Sized Fragments.

AUTHOR(S): Foran, Patrick; Shone, Clifford C.; Dolly, J. Oliver (1)

CORPORATE SOURCE: (1) Dep. Biochem., Imperial Coll., London SW7 2AY UK SOURCE: Biochemistry, (1994) Vol. 33, No. 51, pp. 15365-15374.

ISSN: 0006-2960.

DOCUMENT TYPE: Article LANGUAGE: English

Botulinum neurotoxin serotype B (BoNT/B) and tetanus toxin AΒ (TeTx) block neuroexocytosis through selective endoproteolysis of vesicle-associated membrane protein (VAMP). The enzymological properties of both toxins were compared for the first time in their cleavage of VAMP and various sized fragments using a sensitive chromatographic assay. The optimal substrate sizes for the zinc-dependent protease activities of the light chains of TeTx and BoNT/B were established using synthetic peptides corresponding to the hydrophilic core of VAMP (30-62 amino acids in length). TeTx was found to selectively cleave the largest peptide at a single site, Gln76-Phe77. It exhibited the most demanding specificity, requiring the entire hydrophilic domain (a 62-mer) for notable hydrolysis, whereas BoNT/B efficiently cleaved the much smaller 40-mer. Thus, an unusually long N-terminal sequence of 44 amino acids upstream of the scissile bond is required for the selective hydrolysis of VAMP by TeTx. Using the largest peptide, BoNT/B and TeTx exhibited apprx 50% and 35%, respectively, of the activities shown toward intact VAMP, detergent solubilized from

synaptic vesicles. Given the large size of the smallest substrates, it is possible that these neurotoxins recognize and require a three-dimensional structure. Although both toxins were inactivated by divalent metal chelators, neither was antagonized by phosphoramidon or ASQFETS (a substrate-related peptide that spans the cleavage site), and TeTx was only feebly inhibited by captopril; also, they were distinguishable in their relative activities at different pHs, temperatures, and ionic strengths. These collective findings are important in the design of effective inhibitors for both toxins, as well as in raising the possibility that TeTx and BoNT/B interact somewhat differently with VAMP.

ANSWER 17 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: DOCUMENT NUMBER:

1994:345527 BIOSIS PREV199497358527

TITLE:

A single mutation in the recombinant light chain of tetanus

toxin abolishes its proteolytic activity

and removes the toxicity seen after reconstitution with

native heavy chain.

AUTHOR(S):

Li, Yan; Foran, Patrick; Fairweather, Niel F.; De Paiva, Anton; Weller, Ulrich; Dougan, Gordon; Dolly, J. Oliver (1)

CORPORATE SOURCE: SOURCE:

(1) Dep. Biochem., Imperial Coll., London SW7 2AY UK

Biochemistry, (1994) Vol. 33, No. 22, pp. 7014-7020.

ISSN: 0006-2960.

DOCUMENT TYPE:

Article English

LANGUAGE:

Specific proteolysis by the tetanus toxin light chain of a vesicle-associated membrane protein (VAMP) involved in exocytosis is thought to underlie its intracellular blockade of neurotransmitter release. To substantiate this mechanism, recombinant light chain was expressed as a maltose binding protein-light chain fusion product in Escherichia coli. After purification by affinity chromatography and cleavage with factor Xa, the resultant light chain was isolated and its identity confirmed by Western blotting and N-terminal sequencing. It exhibited activity similar to that of the native light chain in proteolyzing its target in isolated bovine small synaptic vesicles and in hydrolyzing a 62-residue synthetic polypeptide spanning the cleavage site of the substrate. The importance of Glu-234 in the catalytic activity of the light chain, possibly analogous to Glu-143 of thermolysin, was examined using site-directed mutagenesis. Changing Glu-234 to Ala abolished the protease activity of the light chain, but its ability to bind the polypeptide substrate was retained. Each recombinant light chain could be reconstituted with the heavy chain of tetanus toxin, yielding the same level of disulfide-linked species as the two native chains. Whereas the toxin formed with wild-type light chain exhibited appreciable neuromuscular paralysis activity and mouse lethality, the equivalent dichain material containing the Ala-234 mutant lacked neurotoxicity in both the in vitro and in vivo assays. Thus, these results demonstrate directly, for the first time, that the lethality of tetanus toxin and its inhibition of exocytosis in intact neurons are attributable largely, if not exclusively, to endoprotease activity

L8ANSWER 18 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

DUPLICATE 4

ACCESSION NUMBER: 1994:451181 BIOSIS DOCUMENT NUMBER: PREV199497464181

TITLE:

Neuronal involvement in the intestinal effects of

Clostridium difficile toxin A and Vibrio

cholerae enterotoxin in rat ileum.

AUTHOR(S):

Castagliuolo, Ignazio; Lamont, J. Thomas; Letourneau, Richard; Kelly, Ciaran; O'Keane, J. Connor; Jaffer, Amir; Theoharides, Theoharis C.; Pothoulakis, Charalabos (1)

CORPORATE SOURCE: (1) Sect. Gastroenterol., Boston Univ. Med. Center, Univ.

Hosp., 88 East Newton St., Boston, MA 02118 USA

SOURCE: Gastroenterology, (1994) Vol. 107, No. 3, pp. 657-665.

ISSN: 0016-5085.

DOCUMENT TYPE: Article
LANGUAGE: English

Background/Aims: Activation of intestinal mast cells and neurons is involved in intestinal inflammation and diarrhea. This study compared the effects of neuronal inhibitors and inhibition of intestinal sensory afferent nerves on the intestinal actions of Clostridium difficile toxin A, an inflammatory enterotoxin, and cholera toxin, a noninflammatory enterotoxin. Methods: The effects of lidocaine, hexamethonium, atropine, and long-term pretreatment of capsaicin on fluid secretion, mannitol permeability, myeloperoxidase (MPO) activity, and release of rat mast cell protease II (RMCPII) were measured in toxin A- and cholera toxin -exposed loops in vivo. Results: Lidocaine, hexamethonium, and capsaicin, but not atropine, inhibited toxin A-mediated secretion and MPO activity, but only capsaicin reduced mannitol permeability. Lidocaine, but not capsaicin, reduced secretion and permeability caused by cholera toxin. Toxin A caused release of RMCPII from rat ileum in vivo and in vitro; this was inhibited by lidocaine or capsaicin, whereas cholera toxin had no effect on release of RMCPII. Conclusions: Neuronal mechanisms are important in the in vivo effects of these two enterotoxins. Capsaicin-sensitive sensory afferent neurons and mast cells are involved in the intestinal mechanism of toxin A, but not cholera toxin.

L8 ANSWER 19 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1994:227250 BIOSIS DOCUMENT NUMBER: PREV199497240250

TITLE: Antagonism of the intracellular action of botulinum

neurotoxin type A with monoclonal antibodies that map to

light-chain epitopes.

AUTHOR(S): Cenci Di Bello, Isabelle; Poulain, Bernard; Shone, Clifford

C.; Tauc, Ladislav; Dolly, J. Oliver (1)

CORPORATE SOURCE: (1) Dep. Biochem., Imperial Coll. Sci. Technol. and Med.,

South Kensington, London SW7 2AY UK

SOURCE: European Journal of Biochemistry, (1994) Vol. 219, No. 1-2,

pp. 161-169.

ISSN: 0014-2956.

DOCUMENT TYPE: Article LANGUAGE: English

mAbs were produced in mice against highly purified, renatured light chain (LC) of botulinum neurotoxin A (BoNT A) that was immobilised on nitrocellulose to avoid the undesirable use of toxoids. Subcutaneous implants of relatively high amounts (up to 10 mu-g each) of LC allowed its slow release into the systemic circulation and, thus, yielded much higher antibody titres against the underivatized antigen than had hitherto been obtained by conventional immunization. Seven stable hybridoma cell lines were established which secrete mAb of IgG, and IgG-2b subclasses reactive specifically with BoNT A and LC, in native and denatured states, without showing any crossreactivity with types B, E, F or tetanus toxin. The pronounced reactivities of three mAbs towards refolded LC or intact toxin, observed in immunobinding and precipitation assays, relative to that seen in Western blots imply a preference for conformational epitopes. Though mAbs 4, 5 and 7 failed to neutralize the lethality of BoNT in vivo, administration intraneurally of mAb7 prevented the inhibition of transmitter release normally induced by subsequent extracellular administration of BoNT A. Notably, the latter mAb reacted with a synthetic peptide corresponding to amino acids 28-53 in the

N-terminus of the LC, a highly conserved region in Clostridial neurotoxins reported to be essential for maintaining the tertiary structure of the chain. Most importantly, when mAbs 4 or 7 were microinjected inside ganglionic neurons of Aplysia, each reversed, though transiently, the blockade of acetylcholine release by the toxin; this novel finding is discussed in relation to the nature of the zinc-dependent protease activity of the toxin.

ANSWER 20 OF 27 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1993:183439 CAPLUS

DOCUMENT NUMBER: 118:183439

TITLE: Noncytolytic toxin conjugates for

therapeutics

Morgan, Alton Charles, Jr.; Abrams, Paul G. INVENTOR(S):

PATENT ASSIGNEE(S): Neorx Corp., USA

PCT Int. Appl., 72 pp. SOURCE:

CODEN: PIXXD2

DOCUMENT TYPE: Patent English LANGUAGE:

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PATENT NO. KIND DATE APPLICATION NO. DATE PATENT NO. KIND DATE _____ WO 9304191 A1 19930304 WO 1992-US6823 19920813

W: CA, JP

RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, SE PRIORITY APPLN. INFO.: US 1991-745158

Conjugates are disclosed which are useful for modifying target cell functions to achieve therapeutic results. Conjugates may include a noncytolytic toxin that does not directly inhibit protein synthesis and is capable of operating through an existing cellular metab. signalling mechanism conjugated to a targeting moiety that constitutes a ligand recognized by the target cell receptor involved in that existing signalling mechanism. Alternatively, the conjugates may include a toxin domain capable of directly impacting a target cell metabolic process (e.g. catalyzing conversion of ATP to cAMP) or acting on a substrate implicated in such a process (e.g. actin) conjugated with a targeting moiety specific for the target cell population. Methods of using the conjugates are also discussed. In mixed lymphocyte reaction studies with cholera holotoxin and cholera toxin B oligomer, the domain responsible for inhibition of proliferation was assocd. with the toxin's enzymic activity and likely the Al subunit. Construction of a cholera toxin Al/interleukin-2 conjugate for

abrogation of transplant rejection is described, as is modification of retargeted toxin to reduce immunogenicity.

ANSWER 21 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1993:504499 BIOSIS DOCUMENT NUMBER: PREV199396128506

TITLE: Interchange of functional domains switches enzyme

specificity: Construction of a chimeric

pneumococcal-clostridial cell wall lytic enzyme. Croux, C.; Ronda, C.; Lopez, R.; Garcia, J. L. (1)

(1) Unidad Genet. Bacteriana, Cent. Invest. Biol., CORPORATE SOURCE: Velazquez 144, 28006 Madrid Spain

SOURCE: Molecular Microbiology, (1993) Vol. 9, No. 5, pp.

1019-1025.

ISSN: 0950-382X.

DOCUMENT TYPE: Article LANGUAGE: English

AUTHOR(S):

Bacterial autolysins are endogenous enzymes that specifically cleave

covalent bonds in the cell wall. These enzymes show both substrate and bond specificities. The former is related to their interaction with the insoluble substrate whereas the latter determine their site of action. The bond specificity allows their classification as muramidases (lysozymes), glucosaminidases, amidases, and endopeptidases. To demonstrate that the autolysin (LYC muramidase) of Clostridium acetobutylicum ATCC824 presents a domainal organization, a chimeric gene (c/c) containing the regions coding for the catalytic domain of the LYC muramidase and the choline-binding domain of the pneumococcal phage CPL1 muramidase has been constructed by in vitro recombination of the corresponding gene fragments. This chimeric construction codes for a choline-binding protein (CLC) that has been purified using affinity chromatography on DEAE-cellulose. Several biochemical tests demonstrate that this rearrangement of domains has generated an enzyme with a choline-dependent muramidase activity on pneumococcal cell walls. Since the parental LYC muramidase was cholineindependent and unable to degrade pneumococcal cell walls, the formation of this active chimeric enzyme by exchanging protein domains between two enzymes that specifically hydrolyse cell walls of bacteria belonging to different genera shows that a switch on substrate specificity has been achieved. The chimeric CLC muramidase behaved as an autolytic enzyme when it was adsorbed onto a live autolysin-defective mutant of Streptococcus pneumoniae. The construction described here provides experimental support for the theory of modular evolution which assumes that novel proteins have evolved by the assembly of preexisting polypeptide units.

L8 ANSWER 22 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1993:504519 BIOSIS

DOCUMENT NUMBER: PREV199396128526

TITLE: Botulinum neurotoxin A selectively cleaves the synaptic

protein SNAP-25.

AUTHOR(S): Blasi, Juan; Chapman, Edwin R.; Link, Egenharde; Binz,

Thomas; Yamasaki, Shinji; De Camilli, Pietro; Suedhof,

Thomas C.; Niemann, Heiner; Jahn, Reinhard

CORPORATE SOURCE: Howard Hughes Med. Inst., Boyer Center Molecular Med., Yale

Univ. Med. Sch., P.O. Box 9812, New Haven, CT 06536 USA Nature (London), (1993) Vol. 365, No. 6442, pp. 160-163.

ISSN: 0028-0836.

DOCUMENT TYPE: Article LANGUAGE: English

SOURCE:

Neurotransmitter release is potently blocked by a group of structurally related toxin proteins produced by Clostridium botulinum. Botulinum neurotoxin type B (BoNT/B) and tetanus toxin (TeTx) are zinc-dependent proteases that specifically cleave synaptobrevin (VAMP), a membrane protein of synaptic vesicles. Here we report that inhibition of transmitter release from synaptosomes caused by botulinum neurotoxin A (BoNT/A) is associated with the selective proteolysis of the synaptic protein SNAP-25. Furthermore, isolated or recombinant L chain of BoNT/A cleaves SNAP-25 in vitro. Cleavage occurred near the carboxyterminus and was sensitive to divalent cation chelators. In addition, a glutamate residue in the BoNT/A L chain, presumably required to stabilize a water molecule in the zinc-containing catalytic centre, was required for proteolytic activity. These findings demonstrate that BoNT/A acts as a zinc-dependent protease that selectively cleaves SNAP-25. Thus, a second component of the putative

fusion complex mediating synaptic vesicle exocytosis is targeted by a

clostridial neurotoxin.

L8 ANSWER 23 OF 27 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V. on STN

ACCESSION NUMBER: 92174925 EMBASE DOCUMENT NUMBER: 1992174925

TITLE: Proteolytic activity of Clostridium

difficile.

AUTHOR: Seddon S.V.; Borriello S.P.

Microb. Pathogenicity Research Group, MRC Clinical Research CORPORATE SOURCE:

Centre, Watford Road, Harrow, Middlesex HA1 3UJ, United

Kingdom

Journal of Medical Microbiology, (1992) 36/5 (307-311). SOURCE:

ISSN: 0022-2615 CODEN: JMMIAV

United Kingdom_ COUNTRY: DOCUMENT TYPE: Journal; Article 004 Microbiology FILE SEGMENT:

LANGUAGE: English English SUMMARY LANGUAGE:

Ten isolates of Clostridium difficile expressing different degrees of toxigenicity and virulence in an animal model were assayed for the production of proteolytic enzymes by various methods. All strains demonstrated some activity in one or more of the assay systems. There was no direct correlation between toxigenic status and enzyme production. However, those strains known to be highly virulent in a hamster model were the most proteolytic. The most commonly detected enzyme was cell associated and its substrate specificity suggested it was a trypsin-like enzyme. Initial purification of the enzyme from strain VPI 10463 gave a 10% yield with a 14-fold increase in purity. Inhibition studies on this preparation indicated that the enzyme was a thiol protease. The enzyme has pH and temperature optima of 7.5 and 37.degree.C, respectively. These characteristics suggest that the enzyme is more related to clostripain, the thiol clostridio-peptidase of C. histolyticum, than to trypsin. Whilst the role of this enzyme remains unclear, it is possible that it may be a contributory factor in the virulence of the organism as described for other clostridial infections.

ANSWER 24 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

DUPLICATE 5

ACCESSION NUMBER: 1990:49622 BIOSIS

DOCUMENT NUMBER: BA89:26986

INTERRELATIONSHIPS BETWEEN DIGESTIVE PROTEOLYTIC TITLE:

> ACTIVITIES AND PRODUCTION AND QUANTITATION OF TOXINS IN PSEUDOMEMBRANOUS COLITIS INDUCED BY CLOSTRIDIUM-DIFFICILE IN GNOTOBIOTIC MICE.

CORTHIER G; MULLER M C; ELMER G W; LUCAS F; DUBOS-RAMARE F AUTHOR(S):

LABORATOIRE D'ECOLOGIE MICROBIENNE, INSTITUT NATL. DE LA CORPORATE SOURCE:

RECHERCHE AGRONOMIQUE, CRJ, 78350 JOUY-EN-JOSAS, FRANCE.

SOURCE: INFECT IMMUN, (1989) 57 (12), 3922-3927.

CODEN: INFIBR. ISSN: 0019-9567.

BA; OLD FILE SEGMENT:

LANGUAGE: English

Clostridium difficile pathogenicity is related to in vivo production of toxins, and it is of great interest to detect toxins produced in biological samples. Several reports have shown that proteases in stools interfere with immunological methods for quantitation of toxin A. The purpose of this work was to estimate the relationship between the proteases and the C. difficile toxins produced in a gnotobiotic mouse model of pseudomembraneous cecitis. Cecal proteolytic activities hydrolyzed toxin A, and immunoglobulin G bound to the microtiter plate used in immunoassays. This interference could be blocked by the addition of trypsin inhibitor to the samples. The ability of soluble toxin A to bind to bound antibodies in an enzyme-linked immunosorbent assay was not affected by the proteases, but the biological activity was reduced 100-fold. The cytotoxicity of toxin B was not modified by proteolytic activity treatment. Mice inoculated with a low toxin A-producing strain of C. difficile did not die, and no modulation of proteoytic activities occurred. After inoculation with the lethal VPI strain

of C. difficle, toxins A and B reached maximum levels in the ceca at 12 h postinfection. At this time, the proteolytic activities did not decrease from the levels seen at zero time. Mice died within 2 days. At this time (about 32 postinfection), proteolytic activities were sharply decreased in the lower parts of the digestive tracts. The findings that serum inhibited the proteases and that there was a 100-fold increase in-serum-derived mouse immunoglobulins in the lumen as the C. difficle infection progressed suggest that decrease in protease activity in the lower digestive tract may be related to the exudation of serum from the inflammation process.

ANSWER 25 OF 27 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

DUPLICATE 6

1988:51196 BIOSIS ACCESSION NUMBER:

BA85:28055 DOCUMENT NUMBER:

TITLE:

ROLE OF THE ESSENTIAL THIOL GROUP IN THE THIOL-ACTIVATED

CYTOLYSIN FROM CLOSTRIDIUM-PERFRINGENS.

IWAMOTO M; OHNO-IWASHITA Y; ANDO S AUTHOR(S):

DEP. BIOCHEM., TOKYO METROPOLITAN INST. GERONTOL., 35-2 CORPORATE SOURCE:

SAKAECHO, ITABASHI-KU, TOKYO, JPN 173.

EUR J BIOCHEM, (1987) 167 (3), 425-430. SOURCE:

CODEN: EJBCAI. ISSN: 0014-2956.

BA; OLD FILE SEGMENT:

LANGUAGE: English A hemolysin, .theta.-toxin, produced by Clostridium

perfringens has one cysteinyl residue in the free thiol form which is essential for its hemolytic activity. The cysteinyl residue was shown to be located at a position about 5 kDa from the C terminus of the molecule by the method of cysteine-specific chemical cleavage. Modification of the residue with a thiol-blocking agent, 5,5'-dithiobis(2-nitrobenzoic acid), reduced the binding affinity of the toxin to sheep erythrocytes to 1/100 that of intact toxin , resulting in a failure of binding at low cell concentrations (0.5%). Thus the failure of hemolysis at low cell concentrations is primarily ascribed to a decreased affinity of the toxin for erythrocytes. Effects of the modification on the lytic processes were examined using high cell concentrations where considerable amounts of modified toxin bound to the cells. The modified toxin hemolyzes erythrocytes once it binds to them; however, the efficiency of hemolysis is reduced by the modification. These, and additional results indicating that modification alters the sensitivity of toxin molecules to protease digestion, show that thiol-modification inactivates the

toxin by affecting both binding and the subsequent lytic processes, probably though a conformational change introduced in the toxin molecules.

ANSWER 26 OF 27 MEDLINE on STN 77005264 ACCESSION NUMBER: MEDLINE

DOCUMENT NUMBER: 77005264 PubMed ID: 965092

TITLE: Molecular forms of neurotoxins in proteolytic

Clostridium botulinum type B cultures.

AUTHOR: Dasgupta B R; Sugiyama H

SOURCE: INFECTION AND IMMUNITY, (1976 Sep) 14 (3) 680-6.

Journal code: 0246127. ISSN: 0019-9567.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

Priority Journals FILE SEGMENT:

197612 ENTRY MONTH:

ENTRY DATE: Entered STN: 19900313

Last Updated on STN: 19970203

Entered Medline: 19761201

A modified purification method was used to isolate the AB neurotoxin of proteolytic Clostridium botulinum type B strain Lamanna. The preparation was found to be a mixture of two protein forms. They were of molecular weight 152,000 and could not be separated by ion-exchange chromatography or electrophoresis in polyacrylamide gel. was a single polypeptide chain, and the other was a dichain molecule (nicked toxin) held together by an interchain disulfide bond(s). Trypsinization increased the toxicity of the toxin preparation and converted the single-chain molecules into dichain forms that were indistinguishable from the endogenously generated nicked toxin. A protease of the type B culture, with substrate specificity similar to that of trypsin, did not change detectably the molecular form of unnicked type E toxin, although toxicity was increased. Higher toxicity was obtained when unnicked type E was trypsinized; the resulting preparation contained only nicked toxin molecules.

L8 ANSWER 27 OF 27 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1970:53359 CAPLUS

DOCUMENT NUMBER: 72:53359

TITLE: Dynamics of the activity of toxin

components in microbiol matter and culture broths. I

AUTHOR(S): Shemanova, G. F.; Gorshkova, V. I.; Borisova, O. K.;

Shakhanina, K. L.

CORPORATE SOURCE: Inst. Med.-Biol. Probl., Moscow, USSR

SOURCE: Vop. Fiziol. Razmnozheniya Mikroorganizmov Ikh

Identifikatsii (1968), 136-50. Editor(s): Tarkov, M.

I. Izd. "Kartya Moldovenyaske": Kishinev, USSR.

CODEN: 21KZAS

DOCUMENT TYPE: Conference LANGUAGE: Russian

LANGUAGE: Russian

AB Together with the detn. of basic components of toxins

(lecithinase, collagenase, protease) a method was established to det. the activity of toxin components in culture broth isolated from bacterial suspensions of Clostr idium perfringens. A correlation between the secretion max. of collagenase and lecithinase and the active growing state of microbes which stopped completely in 24-36 hr was obsd. The secretion of alk. protease started in 6-9 hr and reached its max. in the 24th hr. In ribosomal fractions and in the hyaloplasm of C. perfringens, alk. protease was found but no lecithinase was detected. Small amts. of collagenase were detd. in the 48-72 hr culture. Decreasing of ribosomes is related to the decreasing of the RNA acid-protein ratio. There was no difference in the sedimentation diagrams of ribosomes coming from the 9 hr culture of C. perfringens toxic or nontoxic strain.

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L9 38 L6 AND TOXIN

=> s 19 and FRET

L10 0 L9 AND FRET

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L11 0 L9 AND FRET

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L12 0 L9 AND FLUOROPHORE

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L13 0 L9 AND ENERGY

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L2 0 S E3 AND CLOSTRIDIUM

L3 0 S E3 AND PROTEASE

L4 1695 S PROTEASE AND CLOSTRIDIUM

L5 873 S L4 AND ACTIVITY

L6 142 S L5 AND METHOD

L7 38 S L6 AND TOXIN

FILE 'STNGUIDE' ENTERED AT 18:06:36 ON 13 SEP 2003

FILE 'BIOSIS, CAPLUS, MEDLINE, WPIDS, EMBASE, SCISEARCH' ENTERED AT 18:09:00 ON 13 SEP 2003

L9 - 38 S L6 AND TOXIN L10 0 S L9 AND FRET L11 0 S L9 AND FRET

L12 0 S L9 AND FLUOROPHORE

L13 0 S L9 AND ENERGY L14 0 S L9 AND STEWARD

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L15 0 L6 AND FRET

=> s 16 and fluorescence

L16 11 L6 AND FLUORESCENCE

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L16 ANSWER 1 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 2003:113075 BIOSIS DOCUMENT NUMBER: PREV200300113075

TITLE: Fluorigenic substrates for the protease

activities of botulinum neurotoxins, serotypes A,

B, and F.

AUTHOR(S): Schmidt, James J. (1); Stafford, Robert G.

CORPORATE SOURCE: (1) Toxinology and Aerobiology Division, United States Army

Medical Research Institute of Infectious Diseases, 1425

Porter St., Fort Detrick, MD, 21702-5011, USA:

james.schmidt@det.amedd.army.mil USA

SOURCE: Applied and Environmental Microbiology, (January 2003,

2003) Vol. 69, No. 1, pp. 297-303. print.

ISSN: 0099-2240.

DOCUMENT TYPE: Article LANGUAGE: English

AB The seven botulinum neurotoxins (BoNTs) are zinc metalloproteases that cleave neuronal proteins involved in neurotransmitter release and are among the most toxic natural products known. High-throughput BoNT assays are needed for use in antibotulinum drug discovery and to characterize BoNT protease activities. Compared to other

BoNT protease activities. Compared to other proteases, BoNTs exhibit unusually stringent substrate requirements with respect to amino acid sequences and polypeptide lengths. Nonetheless, we have devised a strategy for development of fluorigenic BoNT protease assays, based on earlier structure-function studies, that has proven successful for three of the seven serotypes: A, B, and F. In synthetic peptide substrates, the P1 and P3' residues were substituted with 2,4-dinitrophenyl-lysine and S-(N-(4-methyl-7dimethylamino-coumarin-3-yl)-carboxamidomethyl)-cysteine, respectively. By monitoring the BoNT-catalyzed increase in fluorescence over time, initial hydrolysis rates could be obtained in 1 to 2 min when BoNT concentrations were 60 ng/ml (about 1 nM) or higher. Each BoNT cleaved its fluorigenic substrate at the same location as in the neuronal target protein, and kinetic constants indicated that the substrates were selective and efficient. The fluorigenic assay for BoNT B was used to characterize a new competitive inhibitor of BoNT B protease activity with a Ki value of 4 muM. In addition to real-time activity measurements, toxin concentration determinations, and kinetic studies, the BoNT substrates described herein may be directly incorporated into automated high-throughput assay systems to screen large

numbers of compounds for potential antibotulinum drugs.

L16 ANSWER 1 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 2003:113075 BIOSIS DOCUMENT NUMBER: PREV200300113075

TITLE: Fluorigenic substrates for the protease

activities of botulinum neurotoxins, serotypes A,

B, and F.

AUTHOR(S): Schmidt, James J. (1); Stafford, Robert G.

CORPORATE SOURCE: (1) Toxinology and Aerobiology Division, United States Army

Medical Research Institute of Infectious Diseases, 1425

Porter St., Fort Detrick, MD, 21702-5011, USA:

james.schmidt@det.amedd.army.mil USA

SOURCE: Applied and Environmental Microbiology, (January 2003,

2003) Vol. 69, No. 1, pp. 297-303. print.

ISSN: 0099-2240.

DOCUMENT TYPE:

LANGUAGE:

Article English

AB The seven botulinum neurotoxins (BoNTs) are zinc metalloproteases that cleave neuronal proteins involved in neurotransmitter release and are among the most toxic natural products known. High-throughput BoNT assays are needed for use in antibotulinum drug discovery and to characterize

BoNT protease activities. Compared to other proteases, BoNTs exhibit unusually stringent substrate requirements with respect to amino acid sequences and polypeptide lengths. Nonetheless, we have devised a strategy for development of fluorigenic BoNT protease assays, based on earlier structure-function studies, that has proven successful for three of the seven serotypes: A, B, and F. In synthetic peptide substrates, the P1 and P3' residues were substituted with 2,4-dinitrophenyl-lysine and S-(N-(4-methyl-7dimethylamino-coumarin-3-yl)-carboxamidomethyl)-cysteine, respectively. By monitoring the BoNT-catalyzed increase in fluorescence over time, initial hydrolysis rates could be obtained in 1 to 2 min when BoNT concentrations were 60 ng/ml (about 1 nM) or higher. Each BoNT cleaved its fluorigenic substrate at the same location as in the neuronal target protein, and kinetic constants indicated that the substrates were selective and efficient. The fluorigenic assay for BoNT B was used to characterize a new competitive inhibitor of BoNT B protease activity with a Ki value of 4 muM. In addition to real-time activity measurements, toxin concentration determinations, and kinetic studies, the BoNT substrates described herein may be directly incorporated into automated high-throughput assay systems to screen large

L16 ANSWER 2 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 2002:247143 BIOSIS DOCUMENT NUMBER: PREV200200247143

TITLE: Rapid determination of substrate specificity of

Clostridium histolyticum beta-collagenase using an

immobilized peptide library.

numbers of compounds for potential antibotulinum drugs.

AUTHOR(S): Hu, Yongbo; Webb, Erin; Singh, Jasbir; Morgan, Barry A.;

Gainor, James A.; Gordon, Thomas D.; Siahaan, Teruna J. (1) (1) Department of Pharmaceutical Chemistry, University of

CORPORATE SOURCE: (1) Department of Pharmaceutical Chemistry, Univer Kansas, Lawrence, KS, 66045: siahaan@ku.edu USA

SOURCE: Journal of Biological Chemistry, (March 8, 2002) Vol. 277,

No. 10, pp. 8366-8371. http://www.jhc.org/. print.

ISSN: 0021-9258.

DOCUMENT TYPE:

Article

LANGUAGE: English

AB The molecular basis of the substrate specificity of **Clostridium**

histolyticum beta-collagenase was investigated using a combinatorial method. An immobilized positional peptide library, which contains 24,000 sequences, was constructed with a 7-hydroxycoumarin-4-propanoyl (Cop) fluorescent group attached at the N terminus of each sequence. This immobilized peptide library was incubated with C. histolyticum beta-collagenase, releasing fluorogenic fragments in the solution phase. The relative substrate specificity (kcat/Km) for each member of the library was determined by measuring fluorescence intensity in the solution phase. Edman sequencing was used to assign structure to subsites of active substrate mixtures. Collectively, the substrate preference for subsites (P3-P4') of C. histolyticum beta-collagenase was determined. The last position on the C-terminal side in which the identity of the amino acids affects the activity of the enzyme is P4', and an aromatic side chain is preferred in this position. The optimal P1'-P3' extended substrate sequence is P1'-Gly/Ala, P2'-Pro/Xaa, and P3'-Lys/Arg/Pro/Thr/Ser. The Cop group in either the P2 or P3 position is required for a high substrate activity with C. histolyticum beta-collagenase. S2 and S3 sites of the protease play a dominant role in fixing the substrate specificity. The immobilized peptide library proved to be a powerful approach for assessing the substrate specificity of C. histolyticum beta-collagenase, so it may be applied to the study of other proteases of interest.

L16 ANSWER 3 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: DOCUMENT NUMBER:

2001:283768 BIOSIS PREV200100283768

TITLE:

High-throughput fluorogenic assay for determination of

botulinum type B neurotoxin protease

activity.

AUTHOR(S):

Anne, Christine; Cornille, Fabrice; Lenoir, Christine;

Roques, Bernard P. (1)

CORPORATE SOURCE:

(1) Departement de Pharmacochimie Moleculaire et

Structurale, U266 INSERM, UMR 8600 CNRS, UFR des Sciences

Pharmaceutiques et Biologiques, 4, Avenue de

l'Observatoire, 75270, Paris Cedex 06:

roques@pharmacie.univ-paris5.fr France

SOURCE:

Analytical Biochemistry, (April 15, 2001) Vol. 291, No. 2,

pp. 253-261. print.

ISSN: 0003-2697.

DOCUMENT TYPE:

Article English

LANGUAGE: SUMMARY LANGUAGE:

English

Botulinum neurotoxins are responsible for botulism, a flaccid muscular paralysis caused by inhibition of acetylcholine release at the neuromuscular junction. This occurs by cleavage of conserved proteins involved in exocytosis such as synaptobrevin by the zinc metallopeptidase activity of the light chain of some botulinum neurotoxins. Botulism, for which there is presently no therapy available, is a relatively widespread disease that may result in death. Consequently, the development of drugs able to inhibit the hydrolytic activity of these neurotoxins is of great interest. Design and screening of such inhibitors could be largely facilitated by using high-throughput assays. With this aim, a novel in vitro test for quantifying the proteolytic activity of botulinum type B neurotoxin was developed. The substrate is the 60-94 fragment of human synaptobrevin-1 which was modified by introduction of the fluorescent amino acid L-pyrenylalanine in position 74 and a p-nitrophenylalanyl residue as quenching group in position 77. The cleavage of Syb 60-94 (Pya74, Nop77) by the toxin active chain occurs selectively between residues 76 and 77 as in the case of the unmodified synaptobrevin and is directly quantified by measuring the strong fluorescence of the formed metabolite Syb 60-76 (Pya74). This is the easiest, quickest, and cheapest assay described to date for

measuring the proteolytic activity of botulinum type B neurotoxin. It can be easily automated for high-throughput screening. Moreover, amounts of about 3.5 pg/ml of botulinum type B neurotoxin could be detected by this method.

L16 ANSWER 4 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1997:410119 BIOSIS

DOCUMENT NUMBER: PREV199799702162

TITLE: A biotinylated perfringolysin O derivative: A new probe for

detection of cell surface cholesterol.

AUTHOR(S): Iwamoto, Machiko (1); Morita, Ikuo; Fukuda, Mitsugu;

Murota, Sei-Itsu; Ando, Susumu; Ohno-Iwashita, Yoshiko

CORPORATE SOURCE: (1) Dep. Membrane Biochem., Tokyo Metropolitan Inst.

Gerontol., 35-2 Sakae-cho, Itabashi-ku, Tokyo 173 Japan

SOURCE: Biochimica et Biophysica Acta, (1997) Vol. 1327, No. 2, pp.

222-230.

ISSN: 0006-3002.

DOCUMENT TYPE: LANGUAGE:

Article English

theta-Toxin is a cholesterol-binding, pore-forming cytolysin of Clostridium perfringens. To detect cell surface cholesterol, we prepared a theta-toxin derivative, BC-theta by biotinylation of a protease-nicked theta-toxin, which has the same binding affinity for cholesterol as theta-toxin without cytolytic activity. Human erythrocytes, V79 cells and human umbilical vein endothelial cells (HUVEC), were stained with BCO coupled with FITC-avidin, and then the cells were analyzed by either flow cytometry or laser confocal microscopy. The fluorescence intensity increased in both intact and briefly fixed cells when treated with BC-theta. BC-theta-treated V79 cells were stained by neither trypan blue nor propidium iodide, indicating that BC theta stained just the outer surface of the plasma membrane of vital cells. Treatment of the cells with digitonin, a cholesterol sequestering reagent, decreased the fluorescence intensity to the background level, indicating that BC-theta staining is specific for cholesterol. The

fluorescence intensity of erythrocytes pre-permeabilized with a small amount of theta-toxin increased more than ten-fold, suggesting

higher cholesterol contents in the inner layer of the plasma membrane. When cells were cultured with cholesterol-depleted medium, the **fluorescence** intensity stained by BC-theta decreased remarkably in V79 cells, but did not change in HUVEC. This indicates that cell surface cholesterol may be provided in different ways with these two cell lines. These results suggest that BC theta can be a useful probe for visualizing cell surface cholesterol and for evaluating the effects of cellular events

L16 ANSWER 5 OF 11 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC. on STN

ACCESSION NUMBER: 1995:251557 BIOSIS DOCUMENT NUMBER: PREV199598265857

TITLE: Structural studies on the zinc-endopeptidase light chain of

tetanus neurotoxin.

on the topology and distribution of cholesterol.

AUTHOR(S): De Filippis, Vincenzo (1); Vangelista, Luca; Schiavo,

Giampietro; Tonello, Fiorella; Montecucco, Cesare

CORPORATE SOURCE: (1) Centro Ricerca Interdipartimentale Biotechnol.

Innovative, Via Trieste 75, I-35121 Padova Italy

SOURCE: European Journal of Biochemistry, (1995) Vol. 229, No. 1,

pp. 61-69.

ISSN: 0014-2956.

DOCUMENT TYPE: Article LANGUAGE: English

AB Tetanus neurotoxin (TeNT) blocks neuroexocytosis via a zinc-endopeptidase activity highly specific for vescicle-associated membrane

protein (VAMP)/synaptobrevin. TeNT is the prototype of clostridial

neurotoxins, a new family of metalloproteinases. They consist of three domains and the proteolytic activity is displayed by the 50-kDa light chain (L chain). The L chain was isolated here in the native state from bacterial filtrates of Clostridium tetani and its structure was studied via circular dichroism (CD) and fluorescence spectroscopy. The secondary structure content (27% alpha-helix and 43% beta-sheet), estimated by far-ultraviolet CD measurements, was in reasonable agreement with that obtained by standard predictive methods (25% alpha-helix and 49% beta-sheet). Moreover. the hypothetical zinc-binding motif, encompassing residues His-Glu-Leu-Ile-His, was correctly predicted to be in alpha-helical conformation, as also expected on the basis of the geometrical requirements for a correct coordination of the zinc ion. Both near-ultraviolet CD and fluorescence data strongly suggest that the single Trp43 residue is buried and constrained in a hydrophobic environment, likely distant from the zinc ion located in the active-site cleft. The contribution of the bound zinc ion to the overall conformation of TeNT L chain was investigated by different and complementary techniques, including spectroscopic (far- and near-ultraviolet CD, fluorescence, second derivative absorption spectroscopy) as well as proteolytic probes. The results indicate that the zinc ion plays little, if any, role in determining the structural properties of the L chain molecule. Similarly, the metal-free apo-enzyme and the holo-protein share common stability features evaluated in respect to different physico-chemical parameters (pH, temperature and urea concentration). These results parallel those obtained on thermolysin, a zinc-dependent neutral endoprotease from Bacillus thermoproteolyticus, where both conformational and stability properties are unchanged upon zinc removal.

L16 ANSWER 6 OF 11 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER:

2002:218561 CAPLUS

DOCUMENT NUMBER:

137:29736

TITLE:

Rapid determination of substrate specificity of

Clostridium histolyticum .beta.-collagenase

using an immobilized peptide library

AUTHOR(S):

Hu, Yongbo; Webb, Erin; Singh, Jasbir; Morgan, Barry A.; Gainor, James A.; Gordon, Thomas D.; Siahaan,

Teruna J.

CORPORATE SOURCE:

Department of Pharmaceutical Chemistry, The University

of Kansas, Lawrence, KS, 66045, USA

SOURCE:

Journal of Biological Chemistry (2002), 277(10),

8366-8371

CODEN: JBCHA3; ISSN: 0021-9258

PUBLISHER:

American Society for Biochemistry and Molecular

Biology

DOCUMENT TYPE:

Journal English

LANGUAGE: The mol. basis of the substrate specificity of Clostridium AB histolyticum .beta.-collagenase was investigated using a combinatorial method. An immobilized positional peptide library, which contains 24,000 sequences, was constructed with a 7-hydroxycoumarin-4-propanoyl (Cop) fluorescent group attached at the N terminus of each sequence. This immobilized peptide library was incubated with C. histolyticum .beta.-collagenase, releasing fluorogenic fragments in the soln. phase. The relative substrate specificity (kcat/Km) for each member of the library was detd. by measuring fluorescence intensity in the soln. phase. Edman sequencing was used to assign structure to subsites of active substrate mixts. Collectively, the substrate preference for subsites (P3-P4') of C. histolyticum .beta.-collagenase was detd. The last position on the C-terminal side in which the identity of the amino acids affects the activity of the enzyme is P4', and an arom. side chain is preferred in this position. The optimal P1'-P3' extended

substrate sequence is P1'-Gly/Ala, P2'-Pro/Xaa, and P3'-Lys/Arg/Pro/Thr/Ser. The Cop group in either the P2 or P3 position is required for a high substrate activity with C. histolyticum .beta.-collagenase. S2 and S3 sites of the protease play a dominant role in fixing the substrate specificity. The immobilized peptide library proved to be a powerful approach for assessing the substrate specificity of C. histolyticum .beta.-collagenase, so it may be applied to the study of other proteases of interest.

REFERENCE COUNT:

21 THERE ARE 21 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L16 ANSWER 7 OF 11 MEDLINE on STN

ACCESSION NUMBER: 2002139165 MEDLINE

DOCUMENT NUMBER: 21864141 PubMed ID: 11724807

TITLE: Rapid determination of substrate specificity of

Clostridium histolyticum beta-collagenase using an

immobilized peptide library.

AUTHOR: Hu Yongbo; Webb Erin; Singh Jasbir; Morgan Barry A; Gainor

James A; Gordon Thomas D; Siahaan Teruna J

CORPORATE SOURCE: Department of Pharmaceutical Chemistry, the University of

Kansas, Lawrence, Kansas 66045, USA.

SOURCE: JOURNAL OF BIOLOGICAL CHEMISTRY, (2002 Mar 8) 277 (10)

8366-71.

Journal code: 2985121R. ISSN: 0021-9258.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 200204

ENTRY DATE: Entered STN: 20020305

Last Updated on STN: 20030105 Entered Medline: 20020415

The molecular basis of the substrate specificity of Clostridium AΒ histolyticum beta-collagenase was investigated using a combinatorial method. An immobilized positional peptide library, which contains 24,000 sequences, was constructed with a 7-hydroxycoumarin-4-propanoyl (Cop) fluorescent group attached at the N terminus of each sequence. This immobilized peptide library was incubated with C. histolyticum beta-collagenase, releasing fluorogenic fragments in the solution phase. The relative substrate specificity (k(cat)/K(m)) for each member of the library was determined by measuring fluorescence intensity in the solution phase. Edman sequencing was used to assign structure to subsites of active substrate mixtures. Collectively, the substrate preference for subsites (P(3)-P(4)') of C. histolyticum beta-collagenase was determined. The last position on the C-terminal side in which the identity of the amino acids affects the activity of the enzyme is P(4)', and an aromatic side chain is preferred in this position. The optimal P(1)'-P(3)' extended substrate sequence is P(1)'-Gly/Ala, P(2)'-Pro/Xaa, and P(3)'-Lys/Arg/Pro/Thr/Ser. The Cop group in either the P(2) or P(3) position is required for a high substrate activity with C. histolyticum beta-collagenase. S(2) and S(3) sites of the protease play a dominant role in fixing the substrate specificity. The immobilized peptide library proved to be a powerful approach for assessing the substrate specificity of C. histolyticum beta-collagenase, so it may be applied to the study of other proteases of interest.

L16 ANSWER 8 OF 11 MEDLINE on STN ACCESSION NUMBER: 96373022 MEDLINE

DOCUMENT NUMBER: 96373022 PubMed ID: 8776761

TITLE: Expression and purification of a recombinant "small"

sialidase from Clostridium perfringens A99.

AUTHOR: Kruse S; Kleineidam R G; Roggentin P; Schauer R

CORPORATE SOURCE: Biochemisches Institut, Christian-Albrechts-Universitat,

Kiel, Germany.

SOURCE: PROTEIN EXPRESSION AND PURIFICATION, (1996 Jun) 7 (4)

415-22.

Journal code: 9101496. ISSN: 1046-5928.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199612

ENTRY DATE: Entered STN: 19970128

Last Updated on STN: 20000303 Entered Medline: 19961205

A 1.4-kb gene encoding the "small" sialidase isoenzyme of Clostridium perfringens A99, including its own promoter, was AB previously cloned in and expressed by Escherichia coli JM 101. Since all attempts to purify this enzyme to homogeneity were unsuccessful, a new strategy was developed. The structural gene was amplified by means of a PCR technique and inserted into the plasmid vector pQE-10, transferring a six-histidine affinity tag (His6) to the N-terminus of the protein. In order to minimize proteolytic degradation of the sialidase protein, the gene was subcloned into the Escherichia coli strain BL21(DE3)pLys S with reduced protease activity. The sialidase production was increased about 2.5-fold when compared with that of the original clone. The enzyme, released by lysozyme treatment of the bacterial cells, was purified by metal chelate chromatography on Ni-nitrilo-triacetic acid agarose to apparent homogeneity in SDS-PAGE. The 42-kDa protein was enriched 62-fold with a yield of 82% and a specific activity of 280 U mg-1. A total amount of 1 mg sialidase was obtained from 1 liter of bacterial culture. For future studies, including crystallization experiments, the histidine affinity tag was removed from the sialidase enzyme by aminopeptidase K. The sialidase was then separated from aminopeptidase K by ion-exchange chromatography, resulting in an overall yield of 83% and a specific activity of 305 U mg-1 using 4-methylumbelliferyl- alpha-D-N-acetylneuraminic acid under standard conditions. The two forms (with or without the histidine tag) of sialidase exhibited similar kinetic properties when compared to the wild-type enzyme.

L16 ANSWER 9 OF 11 MEDLINE on STN ACCESSION NUMBER: 94071072 MEDLINE

DOCUMENT NUMBER: 94071072 PubMed ID: 8250226

TITLE: Synthesis of N alpha-[3H]acetyl-L-lysine chloromethyl

ketone and its use in the fluorographic detection of

proteases.

AUTHOR: Nishikata M

CORPORATE SOURCE: Central Research Division, School of Dentistry, Hokkaido

University, Sapporo, Japan.

SOURCE: ANALYTICAL BIOCHEMISTRY, (1993 Oct) 214 (1) 222-6.

Journal code: 0370535. ISSN: 0003-2697.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199401

ENTRY DATE: Entered STN: 19940201

Last Updated on STN: 20000303 Entered Medline: 19940105

AB Tritiated N alpha-acetyl-L-lysine chloromethyl ketone (ALCK) was synthesized on a laboratory scale for use as an active-site-directed affinity label in the fluorographic detection of **proteases** after

sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The synthesis involved acetylation of N epsilon-benzyloxycarbonyl-L-lysine chloromethyl ketone with [3H]acetic anhydride just before the removal of the benzyloxycarbonyl group. By this method, [3H]ALCK with a specific activity of 250 mCi/mmol was obtained as a crystal. Trypsin, thrombin, plasmin, papain, and clostripain were inactivated by ALCK according to first-order kinetics. For fluorographic detection of proteases, enzyme samples were allowed to react with [3H]ALCK and then resolved by SDS-PAGE. Proteases that reacted with [3H]ALCK could be detected with a sensitivity equivalent to or higher than that of Coomassie brilliant blue R-250 staining. A trypsin-like protease in Pronase, clostripain as a contaminant in a commercial preparation of Clostridium histolyticum collagenase, and cysteine proteases in Porphyromonas gingivalis could be detected.

L16 ANSWER 10 OF 11 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V. on STN

ACCESSION NUMBER:

2002313187 EMBASE

TITLE:

Rapid determination of substrate specificity of Clostridium histolyticum .beta.-collagenase using

an immobilized peptide library.

AUTHOR:

Hu Y.; Webb E.; Singh J.; Morgan B.A.; Gainor J.A.; Gordon

T.D.; Siahaan T.J.

CORPORATE SOURCE:

T.J. Siahaan, Dept. of Pharmaceutical Chemistry, University

of Kansas, Lawrence, KS 66045, United States.

siahaan@ku.edu

SOURCE:

Journal of Biological Chemistry, (8 Mar 2002) 277/10

(8366-8371).

Refs: 21

ISSN: 0021-9258 CODEN: JBCHA3

COUNTRY:
DOCUMENT TYPE:

United States Journal; Article

FILE SEGMENT:

004 Microbiology 029 Clinical Biochemistry

LANGUAGE: End

English

SUMMARY LANGUAGE:

English

The molecular basis of the substrate specificity of Clostridium histolyticum .beta.-collagenase was investigated using a combinatorial method. An immobilized positional peptide library, which contains 24,000 sequences, was constructed with a 7-hydroxycoumarin-4-propanoyl (Cop) fluorescent group attached at the N terminus of each sequence. This immobilized peptide library was incubated with C. histolyticum .beta.-collagenase, releasing fluorogenic fragments in the solution phase. The relative substrate specificity (k(cat)/K(m)) for each member of the library was determined by measuring fluorescence intensity in the solution phase. Edman sequencing was used to assign structure to subsites of active substrate mixtures. Collectively, the substrate preference for subsites (P(3)-P(4)') of C. histolyticum .beta.-collagenase was determined. The last position on the C-terminal side in which the identity of the amino acids affects the activity of the enzyme is P(4)', and an aromatic side chain is preferred in this position. The optimal P(1)'-P(3)' extended substrate sequence is P(1)'-Gly/Ala, P(2)'-Pro/Xaa, and P(3)'-Lys/Arg/Pro/Thr/Ser. The Cop group in either the P(2) or P(3) position is required for a high substrate activity with C. histolyticum .beta.-collagenase. S(2) and S(3) sites of the protease play a dominant role in fixing the substrate specificity. The immobilized peptide library proved to be a powerful approach for assessing the substrate specificity of C. histolyticum .beta.-collagenase, so it may be applied to the study of other protesses of interest.

L16 ANSWER 11 OF 11 SCISEARCH COPYRIGHT 2003 THOMSON ISI on STN ACCESSION NUMBER: 2002:239678 SCISEARCH

THE GENUINE ARTICLE: 528WR

TITLE: Rapid determination of substrate specificity of

Clostridium histolyticum beta-collagenase using an

immobilized peptide library

Hu Y B; Webb E; Singh J; Morgan B A; Gainor J A; Gordon T AUTHOR:

D; Siahaan T J (Reprint)

-Univ Kansas, Dept Pharmaceut Chem, Lawrence, KS 66045 USA CORPORATE SOURCE:

(Reprint); Sterling Winthrop Res Inst, Collegeville, PA

19426 USA

USA

COUNTRY OF AUTHOR:

SOURCE:

JOURNAL OF BIOLOGICAL CHEMISTRY, (8 MAR 2002) Vol. 277,

No. 10, pp. 8366-8371.

Publisher: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC,

9650 ROCKVILLE PIKE, BETHESDA, MD 20814-3996 USA.

ISSN: 0021-9258.

DOCUMENT TYPE:

Article; Journal

LANGUAGE:

English

REFERENCE COUNT: 21

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

The molecular basis of the substrate specificity of Clostridium AΒ histolyticum beta-collagenase was investigated using a combinatorial method. An immobilized positional peptide library, which contains 24,000 sequences, was constructed with a 7-hydroxycoumarin-4propanoyl (Cop) fluorescent group attached at the N terminus of each sequence. This immobilized peptide library was incubated with C. histolyticum beta-collagenase, releasing fluorogenic fragments in the solution phase. The relative substrate specificity (k(cat)/K-m) for each member of the library was determined by measuring fluorescence intensity in the solution phase. Edman sequencing was used to assign structure to subsites of active substrate mixtures. Collectively, the substrate preference for subsites (P-3-P-4') of C. histolyticum, B-collagenase was determined. The last position on the C-terminal side in which the identity of the amino acids affects the activity of the enzyme is P-4', and an aromatic side chain is preferred in this position. The optimal P-1'-P-3' extended substrate sequence is P-1'-Gly/Ala, P-2'-Pro/Xaa, and P-3'-Lys/Arg/Pro/Thr/Ser. The Cop group in either the P-2 or P-3 position is required for a high substrate activity with C. histolyticum beta-collagenase. S-2 and S. sites of the protease play a dominant role in fixing the substrate specificity. The immobilized peptide library proved to be a powerful approach for assessing the substrate specificity of C histolyticum beta-collagenase, so it may be applied to the study of other proteases of interest.